



TABLE OF CONTENTS

- 1. PURPOSE
- 2. SCOPE
- 3. RESPONSIBILITIES
- 4. REFERENCES
- 5. REAGENTS AND MATERIALS
- 6. EQUIPMENT
- 7. DEFINITIONS
- 8. REGULATORY AGENCIES
- 9. ENVIRONMENTAL RISKS TO RESEARCHERS
- 10. ZOO NOTIC RISKS TO RESEARCHERS
- 11. LAND OWNERSHIP RIGHTS
- 12. TRAINING
- 13. SAFETY PRECAUTIONS
- 14. PERSONAL PROTECTIVE EQUIPMENT
- 15. DECONTAMINATION AND WASTE DISPOSAL
- 16. REMOVAL METHODS
- 17. RELOCATION
- 18. EUTHANASIA AND ACCIDENTAL DEATH
- 19. NECROPSY
- 20. TRANSPORTATION
- 21. PRAIRIE DOG HUSBANDRY
- 22. MARKING ANIMALS FOR IDENTIFICATION
- 23. CONTACT INFORMATION
- 24. LITERATURE CITED

APPENDIX I PLAGUE CASES IN THE U.S.
 APPENDIX II FEVER WATCH FORM
 APPENDIX III SHIPPING MISC. HAZARDOUS MATERIALS
 APPENDIX IV AZGFD SCIENTIFIC COLLECTING LICENSE
 APPENDIX IV AZGFD WILDLIFE SERVICE LICENSE

1. PURPOSE

- a. To describe general regulations for working with prairie dogs in the lab and in the field.
- b. To describe general procedures for providing the highest quality care for prairie dogs, while still allowing the appropriate research or management to be conducted.
- c. To ensure the safety of research personnel.

2. SCOPE

This SOP will cover all basic regulations and procedures for working with prairie dogs (alive or deceased) and prairie dog tissues; both in the lab and in the field. This SOP will be reviewed and updated any time procedures are changed.

3. RESPONSIBILITIES

It is the responsibility of all Northern Arizona University personnel (animal care services, principal investigators, students) to follow these procedures. It is the responsibility of investigators to meet before, during, and after field operations to ensure compliance with this document. It is the responsibility of those who perform work with prairie dogs to review and revise SOPs for appropriateness and accuracy.

4. REFERENCES

Animal Welfare Act (www.aphis.usda.gov/animal_welfare)

Arizona Game and Fish Department (www.azgfd.com)

Centers for Disease Control and Prevention (www.cdc.gov)

Guide for the Care and Use of Laboratory Animals (grants.nih.gov/grants/olaw/Guide-for-the-Care-and-use-of-laboratory-animals.pdf)

Institutional Animal Care and Use Committee (www.aalas.org/iacuc)

5. REAGENTS AND MATERIALS

Note: This list is not intended to cover *all* reagents and materials required for a specific protocol, as many items will be personnel/project specific; nor will every protocol require every item on this list. The purpose of this list is only to serve as a reference for *commonly* used supplies. For more detailed instructions, please reference procedure-specific SOPs.

- a. Biohazard bags (various sizes)
- b. Burlap or cotton sheets (to cover animals to provide sun protection and reduce stress)
- c. Clear packing tape (to tape disposable glove cuffs to sleeves)
- d. Disposable, single-use Tyvek[®] coveralls
- e. Disposable, extended cuff single-use nitrile gloves of the appropriate size
- f. Disposable surgical masks
- g. Euthasol[®] or equivalent
- h. Food/bait (carrots, corn-on-the-cob, apples, sweet feed, sunflower seeds, grass hay)
- i. Formalin
- j. Gallon Ziploc[®] bags (for bagging potentially contaminated clothing worn during field work)
- k. Hand sanitizer or other disinfectant
- l. Identification materials
 - i. Ear tags
 - ii. Passive Integrated Transponder (PIT) tags
 - iii. Hair dye
- m. Insecticide/acaricide dust or spray (e.g., carbaryl, permethrin, DeltaDust[®], OFF![®])
- n. Light colored, fully closed shoes
- o. Light colored long sleeve shirts and pants
- p. Newspaper, hay, or other bedding material
- q. Prescription for appropriate antibiotic prophylactic (e.g., doxycycline)
- r. Respiratory masks (i.e., N95 respirator)
- s. Sharps disposal container
- t. Sunscreen
- u. 1% sodium hypochlorite solution
- v. 10% bleach solution, quaternary ammonium solution, or other EPA registered tuberculocidal disinfectant

6. EQUIPMENT

- a. Digital thermometer
- b. First aid kit

- c. Kevlar® gloves with sleeves
- d. Protective eyewear
- e. Reflective clothing
- f. Rodent nipple drinker or equivalent
- g. Swiffer® Duster
- h. Tomahawk or Havahart live traps
- i. Weighted food bowls

7. DEFINITIONS

BSA – Biological Sciences Annex

Coterie – Family unit; consists of 1 adult male, 2-3 adult females, and all of their young

DOT – Department of Transportation

IACUC – Institutional Animal Care and Use Committee

PPE – Personal Protective Equipment

USDA – United States Department of Agriculture

USGS – United States Geological Survey

8. REGULATORY AGENCIES

United States Department of Agriculture

- a. Prairie dogs are a species regulated by USDA. All NAU policies and procedures follow the guidelines established by USDA.

Institutional Animal Care and Use Committee

- a. The Northern Arizona University Institutional Animal Care and Use Committee oversees compliance with federal regulations regarding use of animals in research or education. To meet these goals, the IACUC requires NAU personnel using animals to meet two obligations:
 - i. Obtain IACUC approval for working with animals via an *Animal Use Protocol Form* <https://nau.edu/Research/Compliance/Animal-Care/Animal-Care-Forms/>
 - ii. Certify that all animal users have undergone appropriate training regarding the regulatory and welfare issues involved in animal use and to become proficient in any animal-related procedures.

Arizona Game and Fish Department

- b. Arizona's native wildlife are protected under Arizona Revised Statutes, Title 17. The Arizona Game and Fish Department includes prairie dogs on the Species of Greatest Conservation Need Tier 1C.
- c. AGFD, under Commission Rule (R12-4-418), mandates that collection/translocation must be approved through:
 - i. A Scientific Collecting License (R12-4-418) (See APPENDIX IV) <https://www.azgfd.com/license/speciallicense/scientificcollection/>
 - a) The license is for the purpose of wildlife management; gathering information valuable to the maintenance of wild populations; the advancement of science; or promotion of the public health or welfare.
 - b) The license is for a purpose that is in the best interest of the wildlife or the species, will not adversely impact other affected wildlife in this state, and may be authorized without posing a threat to wildlife or public safety.
 - c) The license is for a purpose that does not unnecessarily duplicate previously documented projects.
 - d) The applicant has submitted an acceptable proposal as part of the application form.
 - ii. A Wildlife Service License (R12-4-421) (See APPENDIX V)

<https://www.azgfd.com/license/speciallicense/wildlifeservice>

- a) Allows an individual or company to “capture, remove, transport, and relocate to the wild designated live wildlife if the wildlife causes a nuisance, property damage, poses a threat to public health or safety, or if the health or well-being of the wildlife is threatened by its immediate environment”.

9. ENVIRONMENTAL RISKS TO RESEARCHERS

- a. Dehydration
- b. Sunburn
- c. Venomous species (rattlesnakes, spiders, etc.)
- d. Aggressive wildlife (badgers, bears, etc.)

10. ZONOTIC RISKS TO RESEARCHERS

- a. Plague (*Yersinia pestis*) - Plague is an infectious disease caused by the bacterium *Yersinia pestis*. Many species of mammals carry plague, but it is primarily a disease of wild rodents, especially prairie dogs. The bacterium is transmitted by the bites of infected fleas or through contact with infected animals and is a serious concern for both the animals and personnel involved in prairie dog research. Without antibiotic intervention, the mortality rate for pneumonic plague is nearly 100 percent (see www.cdc.gov/plague for more information).

Note: Contact the Coconino County Health Department (<http://www.coconino.az.gov/health>) if a prairie dog study sight shows signs of plague infection (e.g., dead animals above ground, recently active burrows collapsing or becoming overgrown with vegetation, trend of greatly reduced population numbers over time).

- b. Tularemia (*Francisella tularensis*) - Tularemia is an infectious disease caused by the bacterium *Francisella tularensis*. Rabbits, hares, and rodents are especially susceptible and often die in large numbers during outbreaks. The bacterium can be transmitted by the bites of infected ticks or through contact with infected animals (see www.cdc.gov/tularemia for more information).
- c. Rabies – Rabies is a viral disease of mammals, which is most often transmitted through the bite of a rabid animal. This virus infects the central nervous system, ultimately leading to death if untreated (see www.cdc.gov/rabies for more information).
- d. Hantavirus Pulmonary Syndrome – Hantavirus is an infectious disease, which is spread through contact with hantavirus-infected rodents or their urine and droppings (see www.cdc.gov/hantavirus for more information).

11. LAND OWNERSHIP RIGHTS

- a. If a research study site is located on private land, permission must be obtained in writing from the owner of the land *prior* to sample collection.
- b. If a research study site is located on tribal land, permission must first be obtained in writing from the appropriate tribal governing body.
- c. Sampling done near public right-of-ways or on Forest Service/BLM land does not require permission.

12. TRAINING

- a. Occupational Health screening at NAU’s Health and Learning Center (HLC) prior to beginning work with/around prairie dog colonies.
- b. NAU IACUC mandated training, in addition to any other required species-specific or procedure-specific training (e.g., Sharps, basic biosafety).

- i. Zoonotic Risk training (administered by PI or BSA staff)
- ii. Field Safety training (online) and Field Safety form for those doing field work
- c. NAU's Defensive Driving Course if vehicles from NAU Fleet Services are being used (Section 20).

13. SAFETY PRECAUTIONS

- a. Wear procedure appropriate PPE both in the lab and in the field (Section 14)
- b. Antibiotic prophylaxis may be prescribed prior to work when the protocol has a high likelihood of exposure to plague. If this may apply to your work please contact:

Dr. Sandra Smith, MD
Campus Medical Services
(928) 523-2131 (office)
CampusHealth@nau.edu

- c. Tetanus vaccinations are highly recommended for people doing field work.
- d. Do not touch or pick up dead animals if at all possible. Fleas, which may carry plague, jump from carcasses to new hosts.
- e. Minimize contact time with prairie dogs. While some protocols require direct contact with prairie dogs (banding, blood draws, etc.) investigators should make every effort to minimize contact time through attention to preparedness and technique.
- f. All investigators, including students, should practice techniques before handling prairie dogs.
- g. Commercially available insecticides (e.g., DeltaDust®) should be used around prairie dog burrow entrances (if this does not interfere with the focus of the study) to reduce flea burden, and thus reduce exposure to handlers.
- h. After capture, prairie dogs may be dusted or sprayed with an insecticide (e.g., carbaryl, permethrin) to eliminate fleas.
- i. Do not bring companion animals to study sites (i.e., pet dogs), as they can readily become infected with diseases (e.g., plague, rabies), which can be transmitted to humans through respiratory secretions, bites, and scratches.
- j. Clothing should be removed and placed in Ziploc® bags before leaving the test site in order to reduce the likelihood of carrying fleas off-site. Clothing should be laundered before further use.
- k. A fever watch may be necessary when a protocol has a likelihood of plague exposure (see APPENDIX II).
- l. If an occupational exposure occurs (i.e., exposure to prairie dog respiratory secretions, bites or lacerations), seek immediate medical attention. If possible, scrub the affected area with a soft brush and soap for 5 minutes as soon as possible after the incident occurs.
- m. Employ universal sharps precautions (special training may be required):
 - i. The use of sharps should be restricted to situations or procedures where there is no alternative.
 - ii. All persons using sharps must receive appropriate training in the safe use and disposal of sharps.
 - iii. Sharps should never be re-capped, bent, or broken prior to use or disposal.
 - iv. Disposable sharps should be used whenever possible.
 - v. Disposable sharps must be discarded directly and immediately into a sharps disposal container.
 - vi. Disposable sharps are intended for single-use, and should not be re-used.

- vii. Sharps disposal containers must comply with federal and state regulations: leak-proof, puncture-resistant, and have the universal biohazard symbol in red or orange.
- viii. Do not fill sharps containers above the manufacturers marked line.
- ix. For additional disposal information visit <http://nau.edu/research/compliance/environmental-health-and-safety/>

14. PERSONAL PROTECTIVE EQUIPMENT

- a. Gloves: Needed when handling any potentially infectious material
 - i. Kevlar[®] gloves with sleeves: required for the handler during restraint of conscious animals.
 - ii. Disposable, single use double gloves (latex or nitrile): required when performing procedures on restrained, conscious animals.
 - iii. Disposable, single use gloves (latex or nitrile): required any time personnel may be exposed to infectious materials and the above criteria are not met.
- b. Clothing:
 - i. In the field, long pants with tight-fitting socks and long sleeved shirts are recommended. All clothing (including shoes) should be lightly colored for the easy detection of fleas.
 - ii. In the lab, a lab coat or equivalent protection must be worn (reusable or disposable, based on the hazards of the procedure).
- c. Protective eyewear: Needed during procedures that produce splashes or large droplets of blood or other potentially infected fluids.
- d. Surgical Mask: Needed during procedures that produce splashes or large droplets of blood or other potentially infected fluids. Provides mucous membrane protection, but does not provide respiratory protection from infectious aerosols.
- e. Respiratory mask (i.e., N95): Needed when there is a significant risk of aerosolizing zoonotic agents (i.e., during necropsy).
- f. Reflective clothing: Needed when work is performed near roadways, especially at night.
- g. Tyvek[®] coveralls: May be worn as an additional physical barrier when field work is being done in an area with recent plague activity.
- h. Sunscreen: Should be applied as needed when working outside.
- i. Insecticide/acaricide spray (e.g., OFF! [®]): May be applied as a precautionary measure before beginning field work (or immediately after completion of field work when flea collection is the primary goal).
- j. Hand sanitizer or other disinfectant: Should be applied regularly in the field or when hand washing stations are not available

15. DECONTAMINATION AND WASTE DISPOSAL

- a. Waste generated from animals held in captivity (e.g., nesting, food, excrement) should be sprayed 10% bleach solution, quaternary ammonium solution or other EPA registered tuberculocidal disinfectant.
- b. All waste must be double sealed in biohazard plastic bags and disposed of as hazardous waste.
- c. Sharps must be disposed of in leak-proof, puncture-resistant containers marked with the universal biohazard symbol in red or orange; do not fill sharps containers above the manufacturers marked line.
- d. Animal carcasses must be double sealed in biohazard plastic bags and disposed of as hazardous waste. The outer bag should be labeled with protocol information (e.g., date, cage # (if applicable), protocol #, type of animal) and returned to the BSA for freezing and appropriate disposal.

- e. Use cleaning methods which minimize infectious aerosols (i.e., use a Swiffer® Duster in lieu of a broom to avoid stirring dust).
- f. Instruments should be soaked in a freshly prepared 10% bleach solution for a minimum of 10 minutes then rinsed with distilled water after use.
- g. Wash hands after handling animals, bedding material, excreta, etc.

16. REMOVAL METHODS

a. Live-trapping

i. Preferred method (Truett et al. 2001):

- a) Uses Tomahawk or Havahart® metal traps that have a 6-8 inch square door and are 18-24 inches long.
- b) Baited traps are placed at ground level 1-2 meters from active burrows with the entrance of the trap facing the burrow, with a small trail of bait leading from the burrow to the trap.
- c) If time permits, pre-baiting should occur 5-10 days before capture. Pre-baiting is a technique used to accustom the animal to entering the trap, but without trapping. Traps are positioned and baited as described above, but with the door secured in the open position.
- d) After about a week, traps are set for capture (should be done before dawn).
- e) Following capture, no animal should be left in a trap in direct sunlight for more than 10 minutes to avoid dehydration and heat stress.
- f) Animals that must remain in traps for longer than 10 minutes must be provided with shade and moved to a holding cage within 2 hours.
- g) Holding cages will provide sun protection, as well as a constant supply of food and water to minimize stress.

b. Sudsing

Note: Sudsing requires extensive equipment, training and/or professional services, and should only be used when time is of concern (i.e., land development, construction or when other methods have failed). Sudsing increases animal mortality and should never be used as a primary method of removal.

- i. Soapy water is used to flush prairie dogs from their burrows.
- ii. When the prairie dogs exit, they are captured by hand and rinsed with saline to remove soap and dirt from the eyes, then towel dried.
- iii. Animals are then held in family groups in animal kennels or individually in holding cages.
- iv. Holding cages will provide sun protection, as well as a constant supply of food and water to minimize stress.

17. RELOCATION

- a. Prairie dogs hibernate from mid-September through mid-March to late April, when they emerge briefly for mating; about a month later they birth and nurse their young underground (pups emerge mid-June to early July).
- b. The ideal time for translocation is early July to the end of August. This minimizes the potential mortality of juveniles that are still dependent on their mother and, if relocation is the primary goal, allows time for re-habituation (Long et al. 2006).
- c. It is preferable to capture, transport, and release animals within the same day to reduce stress and reduce the likelihood of escape.
- d. Translocations are most successful when coterries are moved together.

- e. If construction is the reason for relocation, it is best to erect temporary barriers at the burrow entrances following the removal of animals to prevent recolonization of the development site.
- f. Affirmation of non-use by other wildlife should be made prior to release at the relocation site.
- g. See Hicks et al. 2015 for detailed relocation procedures.

18. EUTHANASIA AND ACCIDENTAL DEATH

- a. Moribund animals are assessed for treatment by the Attending Veterinarian.
- b. Animals determined to require euthanasia due to injury, disease state, or other factors may be euthanized by an Attending Veterinarian, or their qualified designee, typically by intraperitoneal (IP) injection of Euthasol® (1 mL/10 pounds) or another appropriate euthanasia solution.
- c. Carcasses of euthanized animals should be properly bagged, stored, and disposed of (Section 15d)

19. NECROPSY

- a. Verify death via cervical dislocation or other approved method before further handling.
- b. Employ universal sharps precautions (Section 13.I).
- c. Handle tissue samples with forceps and place into appropriate collection vials.

20. TRANSPORTATION

Note: If an NAU vehicle is being rented, the driver must meet the requirements of NAU's Authorized Driver Program, including passing NAU's Defensive Driving course and quiz, and registering their driver's license. Additional information can be found at <http://nau.edu/facility-services/safety-training/>

a. Live animals

- i. To eliminate the risk of spreading infectious diseases, animals intended for handling or relocation should first be observed to assess overall health. Observation must be done by trained field personnel for field procedures and by the Attending Veterinarian for animals that will be transported to NAU for holding.
- ii. Animals should never be transported or held in temperatures outside of the recommended range of 45-85°F for unadapted mammals (Animal Welfare Act).
- iii. Vehicles must have an air-conditioned cargo hold for animals being transported in temperatures >85°F.
- iv. Animals should be transported in a separate air space from personnel to minimize the risk of transmitting wildlife diseases to humans.
- v. Female and juvenile prairie dogs will tolerate caging with non-family members; adult males only tolerate their coterie-mates and should be placed in individual cages for transport. Special attention should be given so that antagonistic behavior will not occur between animals held in the same cage.
- vi. On the day of capture or transport, animals should be provided with non-perishable chow and water in a weighted bowl or hanging rodent nipple drinker. Cages should be lined with newspaper, hay, or another suitable bedding material.

b. Animal derivatives (e.g., tissue, blood, etc.)

- i. Samples must be transported in compliance with NAU, DOT, and Game and Fish regulations.
- ii. Shipments containing dry ice or formalin must be marked with a Hazard Class 9 label (see APPENDIX III) and are subject to specific shipping regulations and

limitations. Please check with the NAU Environmental Health and Safety to discuss the proper handling of such packages.

- iii. Private vehicles not modified to separate occupant breathing space from the hazardous materials cannot be used for transport.
- iv. Spill kits are strongly recommended for the potential spills of biological agents and chemicals.

21. PRAIRIE DOG HUSBANDRY

a. General

- i. Animals must be held at an IACUC-approved facility, which provides shelter from the elements, protection from encroachment by wild animals, etc.
- ii. To determine the appropriate cage size, please reference the [Guide for the Care and Use of Laboratory Animals](#).
- iii. All cages should provide adequate ventilation and all openings should be appropriately sized to prevent the animals from catching their feet or heads.
- iv. Prairie dogs are social animals and appropriate justification must be made to house them individually.
- v. Female and juvenile prairie dogs will tolerate caging with non-family members; adult males only tolerate their coterie-mates and should be placed in individual cages. Special attention should be given so that antagonistic behavior will not occur between animals held in the same cage.
- vi. To move a prairie dog between cages, place clean cage and soiled cage side-by-side with both doors open, facing each other. Gently nudge prairie dog(s) from behind with a dull stick, encouraging them to move to the clean cage.

b. Daily care

- i. Observe animal daily for signs for poor health (i.e., moribund animals, poor coat quality, hunched posture, inactivity). Report any signs of illness to the Attending Veterinarian.
- ii. Check water bottles. Refill if low.
- iii. Check food hoppers; if low, provide additional rodent chow and low-sodium cattle cake.
- iv. Provide timothy hay and/or clover hay, fresh fruits and vegetables (corn on the cob, baby carrots, apples, lettuce, watermelons, etc.).

c. Every 2-3 days

- i. Change bedding and replace water and food containers.
- ii. With a long handled squeegee or broom, pull dirty bedding to door side of cage for removal. Add new bedding.
- iii. Double-bag waste and place in bin reserved for biohazard waste.

d. Monthly/End of study

- i. Remove prairie dog and place in new clean cage.
- ii. Clean and disinfect all caging and equipment with a 10% bleach solution; rinse and dry.
- iii. Items that can withstand 180°F temperature should be run through cage washer for final cleaning.

22. MARKING ANIMALS FOR IDENTIFICATION

Note: Any of these marking methods can be used, exclusively or in combination, to uniquely mark prairie dogs for subsequent monitoring:

- a. Two uniquely numbered metal ear tags can be affixed on each ear at the base (Monel Small Animal Ear Tag 1005-1; National Band and Tag Co., Newport, KY).

- b. Color-coded ear tags can be used to help identify individuals with binoculars or remote cameras (Nelson and Theimer 2012).
- c. Passive Integrated Transponder (PIT; 134.2 kHz; Biomark, Inc., Boise, ID) can be injected subcutaneously on the back of the neck.
- d. Hair dyes can be used to paint a unique pattern on each animal (Slobodchikoff 1984).

23. CONTACT INFORMATION:

Dr. Sandra Smith, MD
Campus Medical Services
(928) 523-2131 (office)
CampusHealth@nau.edu

Scott Nichols
Attending Veterinarian
(928) 523-7318 (office)
Scott.Nichols@nau.edu

Leslie Hempleman
Animal Care Manager
(928) 523-1330 (office)
(928) 607-9153 (cell)
Leslie.Hempleman@nau.edu

Shelley Jones
Directory of Biological Safety
(928) 523-7268 (office)
Shelley.Jones@nau.edu

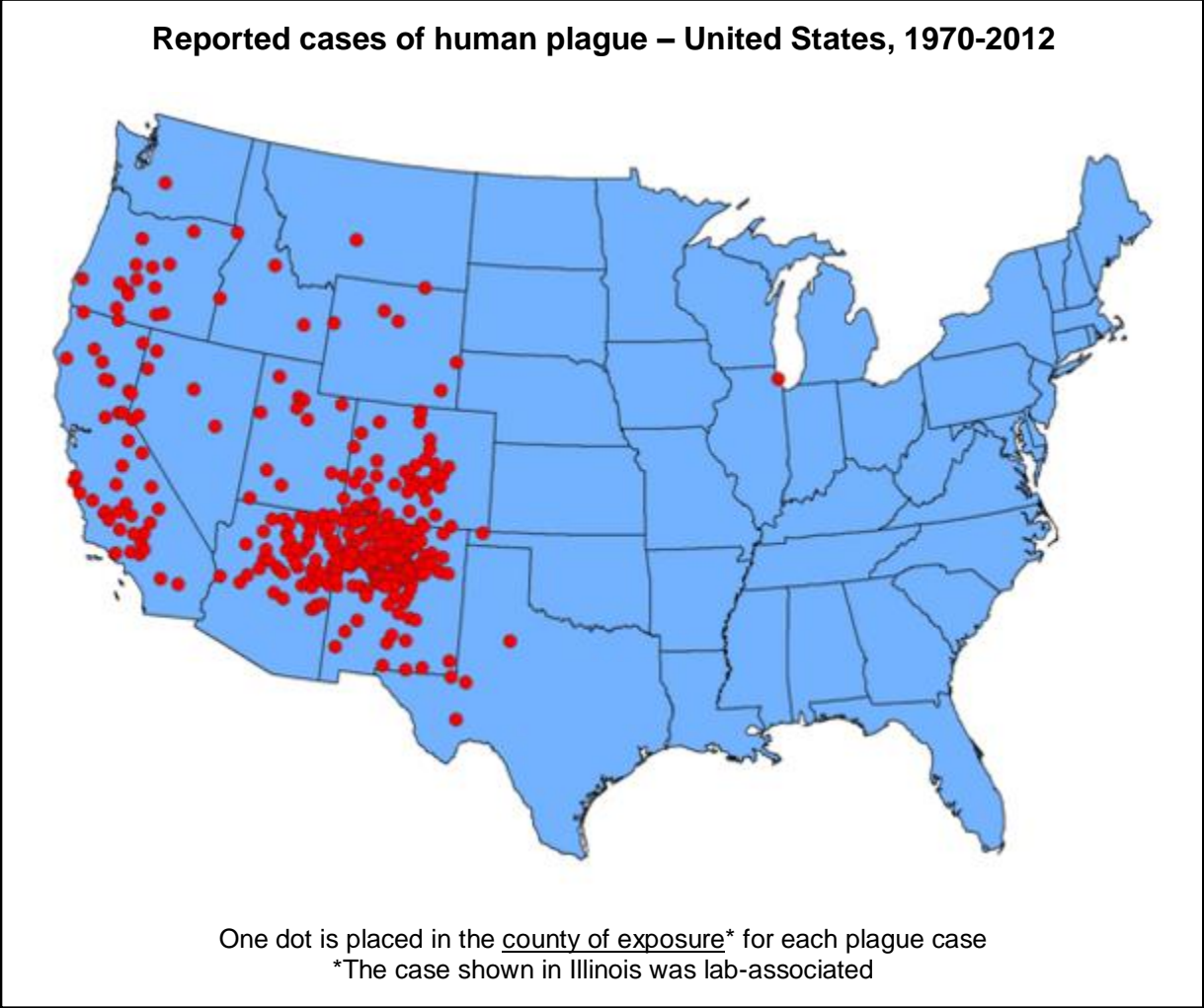
24. LITERATURE CITED

NAU SOP No. AC-09-0030 (Prairie Dog Husbandry)

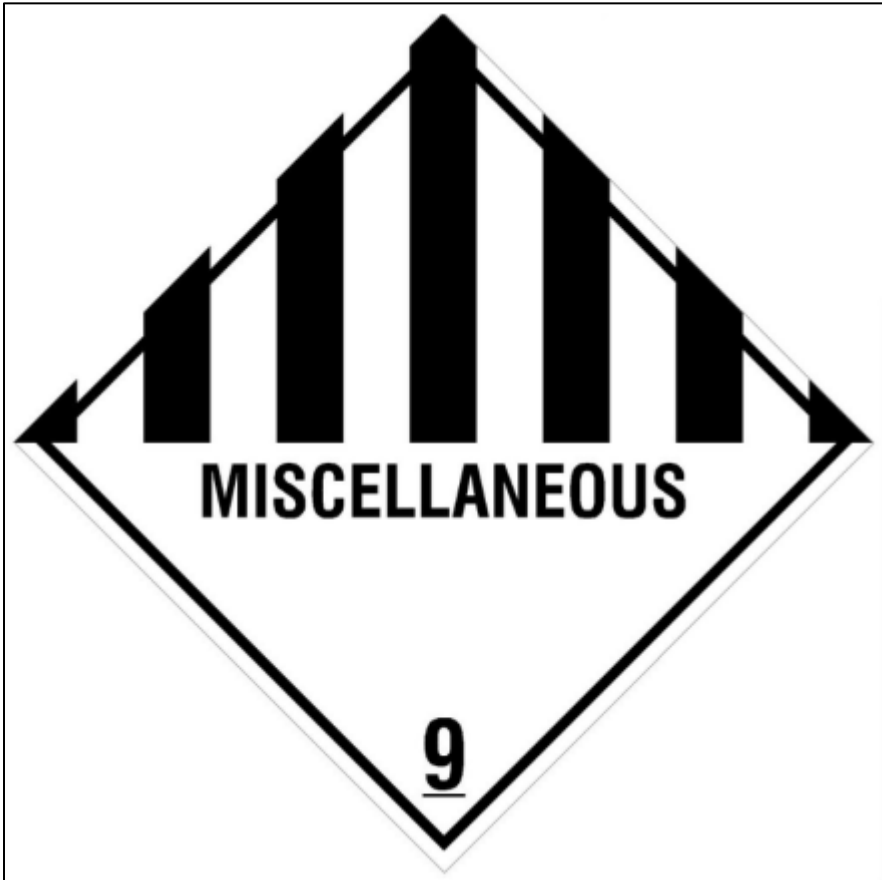
NAU SOP No. AC-09-0029 (Handling of Prairie Dogs)

Hicks, H., J. Cordova, E. Renn. 2015. Translocation Protocol for Gunnison's Prairie Dogs in Arizona. Nongame and Endangered Wildlife Program Technical Report 293. Arizona Game and Fish Department, Phoenix, Arizona.

APPENDIX I



APPENDIX III



APPENDIX IV



ARIZONA GAME & FISH DEPARTMENT
 Terrestrial Wildlife Branch – Scientific Collecting Licenses
 5000 West Carefree Hwy, Phoenix, Arizona 85086

APPLICATION FOR SCIENTIFIC COLLECTING LICENSE- FORM 18A (NO FEE)

CHECK ONE: <input type="checkbox"/> New Applicant <input type="checkbox"/> Renewal		Calendar Year:	
Name:		Phone:	
Male <input type="checkbox"/> Female <input type="checkbox"/>	Birth Date:	Years of Residency (if in AZ):	
Height (ft.-in)	Weight (lbs.)	Eyes:	Hair:
Check all that apply: <input type="checkbox"/> State Agency <input type="checkbox"/> Federal Agency <input type="checkbox"/> Academic Institution <input type="checkbox"/> NGO <input type="checkbox"/> Consulting Firm <input type="checkbox"/> Private <input type="checkbox"/> Other:			
Affiliation:			
Job Title (or nature of affiliation):		Phone (if different than above):	
Affiliation's Address:			
City:	State:	Zip:	
Mailing Address (if different than above):			
City:	State:	Zip:	
Email:	Email (secondary if needed):		

Review Live Wildlife Rule R12-4-409 (General Provisions) and R12-4-418 (Scientific Collecting) prior to applying for a Scientific Collecting License with Arizona Game & Fish Department; available at <https://azgfdportal.az.gov/license/speciallicense/scientificcollection>.

Submit a completed proposal form 18B with application form 18A.

A Scientific Collecting License holder shall submit annual report form 18C (provided) containing the information prescribed therein. Reports are due January 30th following the License calendar year.

If this is a renewal and you still have Arizona wildlife in captivity (in Arizona), you must provide a list of the species and numbers currently in captivity; all such individuals must be placed under a Wildlife Holding License (see Live Wildlife Rule R12-4-417).

By signing this application, I attest that my signature affixed to this application is authentic and the information provided on the application and proposal is true and correct to the best of my knowledge, and my or my agent(s) wildlife privileges are not revoked in this state, any other state, or the United States.

Signature	Date:
-----------	-------

Website: <https://azgfdportal.az.gov/license/speciallicense/scientificcollection/>; Email (preferred): SCPermits@azgfd.gov.

Mail: Arizona Game & Fish Department, Terrestrial Wildlife Branch - SC Licenses, 5000 W Carefree Hwy, Phoenix, AZ 85086; Fax: 623-236-7926. Phone: 623-236-7625

In accordance with provisions under Section 41-1030, Arizona Revised Statutes: An agency shall not base a licensing decision in whole or in part on a licensing requirement or condition that is not specifically authorized by a statute, rule or state tribal gaming compact. A general grant of authority in statute does not constitute a basis for imposing a licensing requirement or condition unless a rule is made pursuant to that general grant of authority that specifically authorizes the requirement or condition. This section may be enforced in a private civil action and relief may be awarded against the state. The court may award reasonable attorney fees, damages and all fees associated with the license application to a party that prevails in an action against the state for a violation of this section. A state employee may not intentionally or knowingly violate this section. A violation of this section is cause for disciplinary action or dismissal pursuant to the Agency's adopted personnel policy. This section does not abrogate the immunity provided by section 12-820.01 or 12-820.02.

FOR DEPARTMENT USE ONLY

Department ID:	License #:	
Date Received:	Date Reviewer Received:	Date Issued:
Reviewer(s) Initials/Date Returned:	# Review Days:	# Total Days:

APPENDIX V

Arizona Game and Fish Department	FOR DEPARTMENT USE ONLY	
	Date Received	Region
	Date Reviewer Received	
	Review Completion Date	
	Approved	
Denied	<u>PLEASE PRINT OR TYPE</u>	
Valid From	To	

****Social Security Number is voluntary-to be used for Sportsman's Database Only**
APPLICATION FOR WILDLIFE SERVICE LICENSE**

FEE: NONE

Name Date of Birth Phone
 Address City State Zip
 Department ID Number/SSN: Email
 Gender Height Weight Eyes Hair
 Business Name
 Address City State Zip
Telephone Numbers: Day **Night**
 Days of the week you will be available for service
 General geographic area of service

List below all species or groups of species of wildlife for which authorization is requested:

List methods of take

Attachments: Additional information required under R12-4-421

Written narrative explaining applicant’s experience in the capture, handling and removal of wildlife, specifying all species for which you have performed such function, referencing the general location and dates that such services were performed, methods used to carry out the activities and methods of disposition for the wildlife captured.

Documentation that proves a minimum of six months full time employment or volunteer experience handling the wildlife listed above.

APPLICANT SIGNATURE: I certify the above is true and correct, and that my live wildlife privileges are not revoked in this or any other state or the United States.

Date _____

Approved By

Date _____

In accordance with provisions under Section 41-1030, Arizona Revised Statutes:

An agency shall not base a licensing decision in whole or in part on a licensing requirement or condition that is not specifically authorized by a statute, rule or state tribal gaming compact. A general grant of authority in statute does not constitute a basis for imposing a licensing requirement or condition unless a rule is made pursuant to that general grant of authority that specifically authorizes the requirement or condition.

This section may be enforced in a private civil action and relief may be awarded against the state. The court may award reasonable attorney fees, damages and all fees associated with the license application to a party that prevails in an action against the state for a violation of this section.

A state employee may not intentionally or knowingly violate this section. A violation of this section is cause for disciplinary action or dismissal pursuant to the Agency’s adopted personnel policy. This section does not abrogate the immunity provided by section 12-820.01 or 12-820.02.